# Corrections

# PHARMACOLOGY

Correction for "Targeting the minor pocket of C5aR for the rational design of an oral allosteric inhibitor for inflammatory and neuropathic pain relief," by Alessio Moriconi, Thiago M. Cunha, Guilherme R. Souza, Alexandre H. Lopes, Fernando Q. Cunha, Victor L. Carneiro, Larissa G. Pinto, Laura Brandolini, Andrea Aramini, Cinzia Bizzarri, Gianluca Bianchini, Andrea R. Beccari, Marco Fanton, Agostino Bruno, Gabriele Costantino, Riccardo Bertini, Emanuela Galliera, Massimo Locati, Sérgio H. Ferreira, Mauro M. Teixeira, and Marcello Allegretti, which appeared in issue 47, November 25, 2014, of *Proc Natl Acad Sci USA* (111:16937–16942; first published November 10, 2014; 10.1073/ pnas.1417365111).

The authors note that an additional affiliation should be listed for Emanuela Galliera. This author's affiliations should appear as "Department of Biomedical, Surgical and Dental Sciences, University of Milan, I-20133 Milan, Italy; and Istituto di Ricerca e Cura a Carattere Scientifico (IRCCS) Galeazzi Orthopaedic Institute, I-20161 Milan, Italy." The corrected author and affiliation lines appear below. The online version has been corrected.

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www.pnas.org/cgi/doi/10.1073/pnas.1422166112

# NEUROSCIENCE

Correction for "Individual differences in the peripheral immune system promote resilience versus susceptibility to social stress," by Georgia E. Hodes, Madeline L. Pfau, Marylene Leboeuf, Sam A. Golden, Daniel J. Christoffel, Dana Bregman, Nicole Rebusi, Mitra Heshmati, Hossein Aleyasin, Brandon L. Warren, Benoit Lebonté, Sarah Horn, Kyle A. Lapidus, Viktoria Stelzhammer, Erik H. F. Wong, Sabine Bahn, Vaishnav Krishnan, Carlos A. Bolaños-Guzman, James W. Murrough, Miriam Merad, and Scott J. Russo, which appeared in issue 45, November 11, 2014 of *Proc Natl Acad Sci* USA (111:16136–16141; first published October 20, 2014; 10.1073/pnas.1415191111).

The authors note that the author name Benoit Lebonté should instead appear as Benoit Labonté. The corrected author line appears below. The online version has been corrected.

Georgia E. Hodes, Madeline L. Pfau, Marylene Leboeuf, Sam A. Golden, Daniel J. Christoffel, Dana Bregman, Nicole Rebusi, Mitra Heshmati, Hossein Aleyasin, Brandon L. Warren, Benoit Labonté, Sarah Horn, Kyle A. Lapidus, Viktoria Stelzhammer, Erik H. F. Wong, Sabine Bahn, Vaishnav Krishnan, Carlos A. Bolaños-Guzman, James W. Murrough, Miriam Merad, and Scott J. Russo

www.pnas.org/cgi/doi/10.1073/pnas.1423575112

# APPLIED PHYSICAL SCIENCES

Correction for "Motionless phase stepping in X-ray phase contrast imaging with a compact source," by Houxun Miao, Lei Chen, Eric E. Bennett, Nick M. Adamo, Andrew A. Gomella, Alexa M. DeLuca, Ajay Patel, Nicole Y. Morgan, and Han Wen, which appeared in issue 48, November 26, 2013, of *Proc Natl Acad Sci USA* (110:19268–19272; first published November 11, 2013; 10.1073/pnas.1311053110).

The authors note that on page 19269, right column, fifth full paragraph, line 4, "200 ms" should instead appear as "200  $\mu$ s."

www.pnas.org/cgi/doi/10.1073/pnas.1423579112

# Motionless phase stepping in X-ray phase contrast imaging with a compact source

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Edited by Robert M. Stroud, University of California, San Francisco, CA, and approved October 21, 2013 (received for review June 10, 2013)

X-ray phase contrast imaging offers a way to visualize the internal structures of an object without the need to deposit significant radiation, and thereby alleviate the main concern in X-ray diagnostic imaging procedures today. Grating-based differential phase contrast imaging techniques are compatible with compact X-ray sources, which is a key requirement for the majority of clinical X-ray modalities. However, these methods are substantially limited by the need for mechanical phase stepping. We describe an electromagnetic phase-stepping method that eliminates mechanical motion, thus removing the constraints in speed, accuracy, and flexibility. The method is broadly applicable to both projection and tomography imaging modes. The transition from mechanical to electromagnetic scanning should greatly facilitate the translation of X-ray phase contrast techniques into mainstream applications.

radiography | CT | retrospective phase stepping | X-ray diffraction | tomosynthesis

oday X-ray imaging modalities account for the majority of the diagnostic imaging procedures in the United States (1). Conventional X-ray images depict variations in the attenuation of transmitted X-rays according to the density distribution in the body. The attenuation-based contrast is typically low in soft tissues at the photon energies commonly used in medical imaging (25-100 keV). However, the phase shift for incident X-rays is many times the linear attenuation of the amplitude in weakly absorbing materials, which points to phase contrast imaging as a means for better resolving soft tissue structures without the need to deposit the harmful radiation (2). Among the first demonstrations of phase contrast in X-ray imaging were diffractionenhanced images using Bragg analyzer crystals (3, 4) and free space propagation of transversely coherent waves (5, 6). In the mid-1990s, monolithic crystal interferometers (7) were used to obtain the first absolute phase shift images of soft tissue samples (2). A few years later, X-ray differential phase contrast (DPC) imaging with a grating interferometer was proposed and then realized (8-10). The grating Talbot interferometer works with a broader energy band and is less affected by environmental changes than monolithic crystal interferometers, at the cost of lower phase sensitivity (11). A major improvement in grating-based imaging was quantitative phase retrieval by the phase-stepping method (10, 12-14). The Talbot-Lau interferometer enabled the use of commercial X-ray tubes for grating DPC, by adding a source grating to give spatial coherence to an extended source (8, 15). A preclinical X-ray DPC computed tomography (CT) scanner incorporating the above advances has been recently demonstrated (16).

Despite the rapid progress in grating-based phase contrast imaging, there are fundamental challenges when it comes to routine applications outside the laboratory environment. Its Achilles' heel is the mechanical phase-stepping process, in which one grating is physically moved in multiple steps over a grating period to obtain a single differential phase image (10, 14). Accurate mechanical movement of centimeter-size objects at the submicron level is inherently slow and difficult to reproduce precisely without a static and stabilized platform. In common configurations including fluoroscopes and CT scanners, the precision motors must be mounted on moving gantries, which will lead to additional mechanical instability (16). Mechanical phase stepping may also be the ultimate limit of the imaging rate, which is critically important for fluoroscopy and clinical CT scans. Recognizing this, two methods for improving the imaging speed in phase contrast CT applications have been reported. One method combines a pair of projection images taken from opposing directions to retrieve phase information, but assumes weak phase objects, no scattering, and a constant, uniform background phase over the whole field of view (17). The second method reduces the number of phase steps by sharing them among neighboring projection angles in a CT scan, but still relies on mechanical phase stepping (18). A more general alternative to phase stepping is the Fourier fringe analysis method, which extracts phase information from a single image containing interference fringes, but with reduced spatial resolution (19-25).

A basic solution to the limitations of mechanical phase stepping should remove the need for physical movement completely. In the fields of radar and ultrasonic imaging, the development of electronic beam steering, which replaced mechanical scanning of the antenna or probe, greatly improved the speed and capability of both technologies (26, 27). We report on an analogous solution for grating-based X-ray phase contrast imaging called electromagnetic phase stepping (EPS). We created an adaptive processing algorithm (28) as part of the method and demonstrate

# **Significance**

From diagnostic exams to security screening, a major concern in X-ray imaging is the potential damage from absorbed radiation energy. Phase contrast techniques are being developed to alleviate the concern by detecting the slight refractive bending of X-rays in an object, instead of relying on the attenuation of the beam. A front runner in the development is technologies that require mechanical scanning of a grating in the X-ray beam to attain high-resolution images. This paper reports a motionless, electromagnetic scanning method in place of mechanical scanning. It lifts the constraints on speed and flexibility and reduces the complexity and cost of the technologies, all of which help bring them closer to everyday applications.

Author contributions: H.W. designed research; H.M., L.C., E.E.B., N.M.A., A.M.D., A.P., and H.W. performed research: H.M. and H.W. contributed new reagents/analytic tools: H.M. and H.W. analyzed data; and H.M., A.A.G., N.Y.M., and H.W. wrote the paper.

Conflict of interest statement: H.W. and H.M. are inventors of a pending patent application of the electromagnetic phase stepping method by the National Institutes of Health. This article is a PNAS Direct Submission

Freely available online through the PNAS open access option.

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This article contains supporting information online at www.pnas.org/lookup/suppl/doi:10. 1073/pnas.1311053110/-/DCSupplemental.

its effectiveness in imaging studies of rodents and other samples in a bench top system.

# Results

A generic grating-based phase contrast imaging system consists of an X-ray tube, a Talbot–Lau interferometer, and an Xray camera as schematically illustrated in Fig. 1*A*. The interferometer has two amplitude gratings ( $G_0$ ,  $G_2$ ) and one phase grating ( $G_1$ ). In our system, the grating period is 4.8 µm. Grating  $G_0$  splits the X-ray cone beam into a number of thin line sources whose lateral coherent lengths are greater than the grating period at the plane of  $G_1$ . Each line source creates an intensity fringe pattern, i.e., fractional Talbot image (10, 29) of  $G_1$ , on the



Fig. 1. Schematic illustration of the electromagnetic phase stepping apparatus and principles. (A) The imaging system (not to scale) is a standard grating interferometer, but with a solenoid coil attached to the X-ray tube housing. The energized coil produces a magnetic field and an associated Lorenz force on the electron beam in the X-ray tube, shifting the impact spot on the anode target from which X-rays are emitted. Correspondingly, the projection image of the sample is displaced in the opposite direction on the camera (displacement exaggerated for better viewing). (B) Schematic representation of electromagnetic phase stepping. The upper red curve represents the moiré fringe pattern, and the lower red curve represents the projection of the object. The red triangle and circle represent a specific location in the projection and its position in the fringe pattern, respectively. In conventional phase stepping (PS), moving one grating shifts the fringe pattern to a new position represented by the dashed blue line. Consequently, the same location in the object projection has a different phase, represented by the blue circle on the dashed blue line. In electromagnetic phase stepping (EPS), the object projection is shifted (dashed green line) due to the displacement of the focal spot of the cone beam, moving the same location in the projection from the red triangle to the green triangle. with a corresponding shift on the fringe pattern (red curve) from the red circle to the green circle. Effectively, the same physical location in the object projection is given a phase shift. (C) Photographs of the home-made solenoid coil and its placement around the cone-beam window of the X-ray tube housing.

plane of  $G_2$ . Because the fringe period is usually smaller than the detector resolution,  $G_2$  is used to produce a broader moiré pattern. When the distance between  $G_0$  and  $G_1$  is the same as that between  $G_1$  and  $G_2$ , the fringe pattern from each individual line source adds up constructively on the plane of  $G_2$ .

If  $G_0$  and  $G_1$  are parallel and  $G_2$  is rotated around the optical axis with respect to  $G_1$  by a small angle  $\theta$ , the differential phase information is encoded into the moiré fringes on the detector plane:

$$I \approx a_0 + a_1 \cos\left[\frac{2\pi}{p}\left(x\theta + \frac{\lambda d}{\pi}\frac{\partial\phi}{\partial y}\right) + \phi_b\right],$$
 [1]

where x and y are coordinates in the detector plane,  $a_0$  is the unmodulated baseline,  $a_1$  is the fringe amplitude, p is the grating period, d is the distance between G<sub>1</sub> and G<sub>2</sub>,  $\lambda$  is the X-ray wavelength, and  $\phi_b$  is the background instrumental phase, which depends on the positions of the gratings. The desired information is the derivative of the X-ray phase shift caused by the sample, expressed as  $\partial \phi / \partial y$  in the detector plane.

The phase-stepping method calculates the differential phase image from several images with different background phases  $\phi_b$ . To date, this has required physically moving one of the gratings in the *y* direction over multiple steps that cover a grating period, while taking an image at each step. In this process, the moiré pattern in the images visibly moves across the static projection of the object, giving rise to the intuitive term of "fringe scanning" as a synonym of phase stepping.

Recognizing that the essential requirement of fringe scanning is a relative movement between the moiré fringes and the projection image of the object, electromagnetic phase stepping achieves the condition by electromagnetically shifting the focal spot of the X-ray tube in a transverse direction across the fringe pattern, e.g., with an externally applied magnetic field that deflects the electron beam in the X-ray tube (Fig. 1A). Shifting the focal spot causes an opposite movement of the projection of the object on the detector plane, while the fringes can be made to remain stationary or move by a different amount. In our setup of the Talbot-Lau interferometer, the moiré fringes are produced by a slight rotation of the third (analyzer) grating. In this case, the fringes remain stationary despite the shifting focal spot. In the inverted embodiment where the moiré fringes are produced by rotating the first (source) grating, the movement of the fringes will exceed that of the projection image. In all cases, the images can then be digitally shifted back to realign the projections of the object. The result is that the fringes move over a stationary projection image, effectively synthesizing the phasestepping process (Fig. 1B).

It is worth noting that shifting the focal spot of the cone beam also causes a slight change of the projection angle on the object. This change is negligible for objects that occupy a small fraction of the distance between the source and the camera. When the object thickness is a significant fraction of the source–camera distance, the reconstruction algorithm takes on the characteristics of stereoscopic imaging or tomosynthesis. A more detailed discussion is provided in *SI Text*.

In our imaging device, the gratings are rigidly mounted. A solenoid coil is attached to the front surface of the X-ray source (Fig. 1*C*) and is driven by a 25-V power supply to produce the magnetic field with full digital control and a response time of 200 ms. The resulting focal spot shift causes an opposite movement of the object projection on the camera over a stationary fringe pattern (Movie S1). A magnetic field of 2.4 mT was sufficient to shift the projected image over one period of the moiré fringes (300  $\mu$ m). The details of the experimental setup are described in *Methods*.

We created an adaptive image-processing algorithm to extract the differential phase contrast, scatter (dark-field) (20, 30), and conventional attenuation images from the EPS data without prior knowledge of the movement of the object projection or of the moiré fringes (*SI Text*). The algorithm first aligns the projections of the object, generating an image stack in which the object is stationary while the moiré fringe pattern moves over it. The aligned images are then processed as fringe-scanned images by the second part of the algorithm, which measures the phase increments in the phase-stepping process with a Fourier transform method (2, 19, 20, 31). The algorithm can cope with arbitrary and spatially varying phase increments without assuming a priori knowledge, and thus is robust against potential instabilities in the alignment of various components in the imaging system.

Using the bench top system with electromagnetic phase stepping, we first imaged a reference sample containing borosilicate spheres of 5-mm diameter. The full series of images is included in Movie S1. Fig. 2 shows the processed DPC, absolute phase shift (via direct integration of the DPC), and linear intensity attenuation images. The stripe artifacts in the phase shift image result from the lack of low-spatial frequency information in the DPC signal. The linear attenuation at the sphere centers is  $0.61 \pm 0.02$ , corresponding to an effective mean X-ray energy of  $35.5 \pm 0.5$  keV. The total phase shift at the sphere centers is estimated to be  $(3.2 \pm 0.2) \times 10^2$  rad, from which the real part of the refractive index decrement of the borosilicate material is estimated to be  $\delta = (3.6 \pm 0.3) \times 10^{-7}$ , slightly below the tabulated reference value of  $3.7 \times 10^{-7}$  for 35.5-keV photon energy (http://physics.nist.gov/PhysRefData/FFast/html/form.html).

As an example of a biological specimen, Fig. 3 shows the DPC and linear attenuation images of a cricket obtained by electromagnetic phase stepping. The DPC image reveals more detailed



**Fig. 2.** Images of a reference sample containing borosilicate spheres. (A) Differential phase contrast, (*B*) phase shift, obtained by direct integration of the DPC information and baseline corrected through linear fitting, (*C*) linear intensity attenuation, and (*D*) a cross-section profile of the phase shift through the center of a sphere, the location of which is marked by the green line in *B*. The blue curve is the experimental data, and the red curve is a fit of the profile of the spheres to the data, from which an estimate of the refractive index decrement could be made. This was  $(3.6 \pm 0.3) \times 10^{-7}$ . The effective mean photon energy was estimated to be 35.5 keV from the level of intensity attenuation through the spheres in *C*. At this energy, the refreere value of refractive index decrement for borosilicate is  $3.7 \times 10^{-7}$  (http://physics.nist.gov/PhysRefData/FFast/html/form.html), which is slightly above the estimate from the phase shift in *D*.



**Fig. 3.** Reconstructed images of a cricket. (*A*) Linear intensity attenuation; (*B*) differential phase contrast. A tungsten bead of 0.8-mm diameter (bright dot in *A*) was placed near the head of the cricket as a marker. Such markers are used to accurately determine the displacement of the projection images during electromagnetic phase stepping.

structures throughout the head, the body, and the legs of the cricket, owing to its sensitive nature to small changes in the density of the constituents.

Biomedical research routinely involves the imaging of rodents, and the photon energy of the bench top system was sufficient to penetrate the body of adult mice. As examples, under a protocol approved by the Animal Care and Use Committee of the National Heart, Lung, and Blood Institute, we imaged the head region of a euthanized mouse in air and the torso region of another euthanized mouse fixed in 10% (vol/vol) buffered formalin.

Fig. 4 A-D are the processed images of the head region, including the DPC, phase contrast-enhanced, linear intensity attenuation, and scatter (dark-field) linear extinction images. The DPC signal degrades into random phase noise in the areas of the metallic ear tag due to the strong attenuation of the fringes. Phase retrieval by direct integration of the DPC image can lead to substantial errors for reasons. The first relates to the inherent lack of low-frequency information in the DPC data, and the second factor is the random phase values in areas where the interference fringes are strongly suppressed, either due to attenuation (as around the metallic ear tag) or scattering. The first can be addressed by the method of Roessl et al. (32), which merges the low-spatial frequency content of the intensity attenuation with the high-spatial frequency content of the DPC data according to the scaling between the real and imaginary parts of the refractive index of the material (here, soft tissue). The second problem is circumvented by substituting the derivative of the intensity attenuation into the DPC image in areas where the DPC information is missing, again using an appropriate scaling factor. These calculations are described in some detail in Methods. The end result is a phase contrast-enhanced image (Fig. 4B). As indicated by the white arrows, numerous details emerge in the phase contrast-enhanced image that cannot be clearly seen in the intensity attenuation image.

Fig. 5 is a compilation of reconstructed images of the torso region of the mouse. The DPC image highlights weakly absorbing phase objects such as air bubbles (Fig. 5*A*). The phase contrastenhanced image (Fig. 5*B*) contains both the phase and attenuation information. The lungs are most visible in the dark-field (scatter) image (Fig. 5*C*), owing to their porous microstructures. The highdensity bones and the metallic ear tag are clearly visible in the intensity attenuation image (Fig. 5*D*).

Movies of the mouse head (Movie S2) and torso images (Movie S3) are provided for comparison between phase contrastenhanced and linear intensity attenuation images.



**Fig. 4.** Reconstructed images of the head region of a mouse. (*A*) Differential phase contrast, (*B*) phase contrast enhanced, (*C*) dark field (scatter), and (*D*) linear intensity attenuation. The arrows in *B* indicate examples of features more visible in the phase contrast-enhanced image than in the classic intensity attenuation image (*D*). The bright U-shaped object is a metallic ear tag.

### Discussion

Grating interferometers used with phase stepping enable highresolution X-ray phase contrast imaging with compact X-ray tubes. However, the stringent requirements for mechanical phase stepping have been a major challenge in bringing phase contrast into common imaging systems. The electromagnetic phase-stepping method and the adaptive processing algorithms presented here effectively replace the precision mechanical scanning system and its associated engineering challenges with a simple solenoid coil attached to the X-ray source, providing substantial advantages in speed, accuracy, and flexibility. The near instantaneous control of the focal spot could also enable real-time compensation for instrumental instabilities, including thermal drift and vibrations. The transition from mechanical to electromagnetic scanning also reduces the cost of parts and maintenance and should improve reliability, all of which may contribute to the translation of phase contrast techniques into mainstream applications. For biomedical imaging, grating periods of a few hundred nanometers are being developed for greater phase sensitivity (33, 34). Here, electromagnetic phase stepping may become a necessity, as precise mechanical movement at the nanometric level may be difficult to achieve outside the most favorable settings.

### Methods

Technical Specifications of the Imaging System. We used a tungsten-target X-ray tube (SB-80-1k; Source Ray) operating at a peak voltage of 55 kV and a current of 1 mA as the source. The focal spot of the tube was ~50  $\mu$ m. The



For electromagnetic phase stepping, a home-made solenoid coil of copper wire (60-mm inner diameter, 200 turns) was attached to the front surface of the X-ray tube housing (Fig. 1*B*). The coil was driven by a digital power supply which provided up to 2.0 A of current at up to 8 W of power. The corresponding peak magnetic field was calculated to be 3.1 mT at the location of the electron beam inside the X-ray tube. The field strength was verified experimentally with a magnetometer. The electron beam is oriented vertically. The magnetic field shifted the focal spot by up to 380  $\mu$ m (with 1.5-A current applied) in the horizontal direction, perpendicular to the moiré fringes. The deflections of the focal spot at various levels of input current into the coil were measured experimentally as shown in Fig. S1.

**Image Acquisition and Processing in Electromagnetic Phase Stepping.** The method of electromagnetic phase stepping obtains three types of information



**Fig. 5.** Reconstructed images of the torso region of a mouse. (*A*) Differential phase contrast, (*B*) phase contrast enhanced, (*C*) dark field (scatter), and (*D*) linear intensity attenuation. The lungs are most clearly seen in the scattering image (*C*).

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from a single set of raw images: the differential phase, the conventional linear attenuation, and the dark-field (scatter) images. For the method to be broadly applicable, the image reconstruction algorithm needs to be able to cope with a number of instabilities that may be present in a compact imaging device. These include drift of the focal spot of the X-ray tube, drift in the alignment of the gratings and other components, and variable positioning of the imaged object.

Each phase-stepping cycle includes six progressive levels of input current into the field coil from 0 to 1.5 A, resulting in six different positions of the cone beam focal spot (Fig. 1A). At each position, an image is taken. The image contains a moiré fringe pattern, which is modulated in amplitude and in phase by the projection of the object. A shift of the focal spot results in a displacement of the projection in the opposite direction, as well as a change of the projection angle. The moiré fringes remain stationary with the specific arrangement of the gratings in our device described above. If we make the assumption that the thickness of the object takes up a small fraction of the distance between the source and the camera, the movement of the projection is approximately uniform throughout the sample, and the change in projection angle can be neglected. More generally, the movement of the projection of a given transverse plane across the beam axis (focal plane) is determined by its position between the source and the detector. Therefore, the image reconstruction effectively focuses on a transverse slice through the object. This is similar to tomosynthesis.

The reconstruction algorithm is adaptive in two ways: first, owing to variable sample positioning, the movement of the projection of the object is not assumed in advance but determined retrospectively from the images themselves; and second, once the projections are aligned, the algorithm needs to retrieve phase and amplitude images from a set of arbitrary fringe positions, i.e., nonuniform and spatially varying phase increments between successive images in a phase-stepping set.

The displacement of the sample projection on the detector plane is measured through a Fourier space analysis that demodulates the moiré fringes while retaining the projection image at a reduced resolution (2, 19, 20). The relative movement of the projections is measured from the demodulated

- Smith-Bindman R, et al. (2012) Use of diagnostic imaging studies and associated radiation exposure for patients enrolled in large integrated health care systems, 1996– 2010. JAMA 307(22):2400–2409.
- Momose A, Fukuda J (1995) Phase-contrast radiographs of nonstained rat cerebellar specimen. *Med Phys* 22(4):375–379.
- Davis TJ, Gao D, Gureyev TE, Stevenson AW, Wilkins SW (1995) Phase-contrast imaging of weakly absorbing materials using hard x-rays. *Nature* 373(6515):595–598.
- Chapman D, et al. (1997) Diffraction enhanced x-ray imaging. *Phys Med Biol* 42(11): 2015–2025.
- Snigirev A, Snigireva I, Kohn V, Kuznetsov S, Schelokov I (1995) On the possibilities of x-ray phase contrast microimaging by coherent high-energy synchrotron radiation. *Rev Sci Instrum* 66(12):5486–5492.
- Wilkins SW, Gureyev TE, Gao D, Pogany A, Stevenson AW (1996) Phase-contrast imaging using polychromatic hard X-rays. Nature 384(6607):335–338.
- 7. Bonse U, Hart M (1965) An X-ray interferometer. Appl Phys Lett 6(8):155.
- 8. Clauser JF (1998) US Patent 5,812,629.
- David C, Nohammer B, Solak HH, Ziegler E (2002) Differential x-ray phase contrast imaging using a shearing interferometer. Appl Phys Lett 81(17):3287–3289.
- Momose A, et al. (2003) Demonstration of X-ray Talbot interferometry. Jpn J Appl Phys 42(Pt 2, 7B):L866–L868.
- Momose A (2003) Phase-sensitive imaging and phase tomography using X-ray interferometers. Opt Express 11(19):2303–2314.
- Bruning JH, et al. (1974) Digital wavefront measuring interferometer for testing optical surfaces and lenses. Appl Opt 13(11):2693–2703.
- Momose A (1995) Demonstration of phase-contrast x-ray computed-tomography using an x-ray interferometer. Nucl Instrum Methods Phys Res A 352(3):622–628.
- Weitkamp T, et al. (2005) X-ray phase imaging with a grating interferometer. Opt Express 13(16):6296–6304.
- Pfeiffer F, Weitkamp T, Bunk O, David C (2006) Phase retrieval and differential phasecontrast imaging with low-brilliance X-ray sources. Nat Phys 2(4):258–261.
- Tapfer A, et al. (2012) Experimental results from a preclinical X-ray phase-contrast CT scanner. Proc Natl Acad Sci USA 109(39):15691–15696.
- Zhu P, et al. (2010) Low-dose, simple, and fast grating-based X-ray phase-contrast imaging. Proc Natl Acad Sci USA 107(31):13576–13581.
- Zanette I, et al. (2012) Trimodal low-dose X-ray tomography. Proc Natl Acad Sci USA 109(26):10199–10204.
- Takeda M, Ina H, Kobayashi S (1982) Fourier-transform method of fringe-pattern analysis for computer-based topography and Interferometry. J Opt Soc Am 72(1): 156–160.
- Wen H, Bennett EE, Hegedus MM, Carroll SC (2008) Spatial harmonic imaging of X-ray scattering—initial results. *IEEE Trans Med Imaging* 27(8):997–1002.

images. The full-resolution images are then shifted by the opposite amounts to align all of the projections. In the aligned images, the projection of the object remains static while the moiré fringes move across it. The intensity at each pixel oscillates with the moving fringes. In effect, the aligned images are equivalent to images acquired in mechanical phase stepping by moving one of the gratings. The details of the reconstruction procedure after the alignment step are described in *SI Text*.

Reconstruction of Phase Contrast-Enhanced Images. The algorithm is as follows: if we define  $A_0$  and  $\phi$  as the linear attenuation and the phase shift of the X-ray wave front after propagation through the object,  $A_0$  is simply the absolute value of the natural logarithm of the transmission, and  $\frac{\partial d}{\partial v}$ corresponds to the DPC signal. The first step is to incorporate the derivative of the linear attenuation  $\frac{\partial A_0}{\partial y}$  into the DPC signal in a weighted sum of  $\frac{\partial A_c}{\partial v} = CW_0 \frac{\partial A_0}{\partial v} + W_1 \frac{\partial \Phi}{\partial v}$ , where C is the scaling factor between the real and imaginary parts of the refractive index (32), and the weights  $W_0$  and  $W_1$ are determined locally according to the amplitude of the interference fringes  $A_1$  and the noise level  $N_1$  in the fringe amplitudes. Specifically,  $W_0(r) = 1/\{1 + [A_1(r)/(2N_1)]^6\}$ , and  $W_1(r) = 1 - W_0(r)$ , Once the combined differential image  $\frac{\partial A_c}{\partial y}$  is determined, it is merged with the intensity attenuation data into a phase contrast enhanced image according to the algorithm described in ref. 31: the direct integral of  $\frac{\partial A_c}{\partial u}$  is high-pass filtered in the Fourier space, and merged with the low-spatial frequency part of the linear intensity attenuation, and then inverse Fourier transformed into the final image.

ACKNOWLEDGMENTS. We are grateful to the staff of the Nanofab Facility of National Institute of Standards and Technology for assistance with fabrication of the phase grating, and to Prof. Marco Stampanoni of Paul Scherrer Institute for helpful discussion on the fusion of phase and intensity images. This work was supported by the Intramural Research Program of the National Institutes of Health, including the National Heart, Lung, and Blood Institute and the National Institute of Biomedical Imaging and Bioengineering.

- Wen H, Bennett EE, Hegedus MM, Rapacchi S (2009) Fourier X-ray scattering radiography yields bone structural information. *Radiology* 251(3):910–918.
- Momose A, Yashiro W, Maikusa H, Takeda Y (2009) High-speed X-ray phase imaging and X-ray phase tomography with Talbot interferometer and white synchrotron radiation. Opt Express 17(15):12540–12545.
- Bennett EE, Kopace R, Stein AF, Wen H (2010) A grating-based single-shot x-ray phase contrast and diffraction method for in vivo imaging. *Med Phys* 37(11):6047–6054.
- Sato G, et al. (2011) Two-dimensional gratings-based phase-contrast imaging using a conventional x-ray tube. Opt Lett 36(18):3551–3553.
- Morgan KS, Paganin DM, Siu KKW (2011) Quantitative x-ray phase-contrast imaging using a single grating of comparable pitch to sample feature size. Opt Lett 36(1): 55–57.
- Fowler CA (1998) Old radar types never die; they just phased array or ... 55 years of trying to avoid mechanical scan. *IEEE Aerosp Elect Syst Mag* 13(9):24A–24L.
- Thomenius KE (2003) Instrumentation Design for Ultrosonic Imaging (McGraw-Hill, New York).
- Wen H, Miao H, Bennett EE, Adamo NM, Chen L (2013) Flexible retrospective phase stepping in X-ray scatter correction and phase contrast imaging using structured illumination. *PLoS One* 8(10):e78276.
- Cloetens P, Guigay JP, De Martino C, Baruchel J, Schlenker M (1997) Fractional Talbot imaging of phase gratings with hard x rays. Opt Lett 22(14):1059–1061.
- Pfeiffer F, et al. (2008) Hard-X-ray dark-field imaging using a grating interferometer. Nat Mater 7(2):134–137.
- Goldberg KA, Bokor J (2001) Fourier-transform method of phase-shift determination. Appl Opt 40(17):2886–2894.
- Roessl E, et al. (2012) Image fusion algorithm for differential phase contrast imaging. Proc SPIE 8313:831354.
- Lynch SK, et al. (2012) Fabrication of 200 nanometer period centimeter area hard x-ray absorption gratings by multilayer deposition. J Micromech Microeng 22(10): 105007.
- Wen H, et al. (2013) Interferometric hard x-ray phase contrast imaging at 204 nm grating period. Rev Sci Instrum 84(1):013706.
- Reznikova E, Mohr J, Boerner M, Nazmov V, Jakobs PJ (2008) Soft X-ray lithography of high aspect ratio SU8 submicron structures. (Translated from English). *Microsyst Technol* 14(9-11):1683–1688.
- Noda D, et al. (2008) Fabrication of large area diffraction grating using LIGA process. Microsyst Technol 14(9-11):1311–1315.
- Hoshino M, Uesugi K, Yagi N (2012) Phase-contrast X-ray microtomography of mouse fetus. Biol Open 1(3):269–274.
- Wu B, Kumar A, Pamarthy S (2010) High aspect ratio silicon etch: A review. J Appl Phys 108(5):051101.

